

# Interior resistance programs

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# Breeding for resistance programs - considerations

## Impacts:

- Biological
  - Pest
  - Host
- Ecological/Environmental:
  - Role of host, other species affected, sensitive habitat
- Social – recreational, public/stakeholder concern
- Cultural - host or pest may have cultural significance
- Economic



# Breeding for resistance programs - considerations

Demand

Feasibility of success

Cost of resistance

Consequence of no action



# Pest resistance screening strategies

## Field-based

- more parents evaluated
- longer time (10 years)
- easier to link to growth on landscape
- no guarantee proper disease incidence
- less work



## Artificial

- fewer parents evaluated
- shorter time (3-4 years)
- harder to link to growth on landscape
- good homogeneous infection
- labour intensive



# Considerations for establishment of an artificial inoculation and screening program

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## Obtaining the pest/pathogen

- Collecting from the environment
- Obtaining from partners
- Long term storage, rearing, maintenance of cultures/pests



# Considerations for establishment of an artificial inoculation and screening program

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Propagating host material for screening:

- Collecting seed/scion from environment
- Propagating
- Obtaining from partners





# Considerations for establishment of an artificial inoculation and screening program

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- Applying pest/pathogen to host:
  - What propagule/form/life stage do we use
  - How much is needed for disease?
  - What method do we use to apply?

# Considerations for establish an artificial inoculation and screening program

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- Post inoculation/infestation treatment and storage?
- Experimental design
- Post screening objectives
- Funding
- Personnel
- Facilities





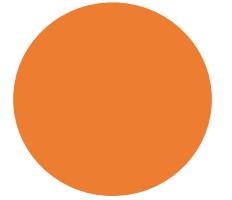
# Current programs

- *Armillaria ostoyae* (Fdi)
  - Artificial inoculations and progeny trials
- *Pissodes strobi* (Sx)
  - Augmentation and field trials
  - Proposed in 2024: Chem ecology – host/pest interactions
- *Dothistroma septosprum* (Pli)
  - Artificial inoculations and progeny trials
- *Cronartium ribicola* (Pa & Pw)
  - Artificial inoculations and progeny trials



# Additions in 2024

- Proposed *Phytophthora spp.* root rot (Pw)
  - Product efficacy trial, collaboration with orchard colleagues. Likely to start 2025 or 2026.
- *Cronartium comandrae* (Pli)
  - Planting new site and future site selections
- *Endocronartium harkenessii* (Py)
  - Isolations and challenge method develop.



# Fdi *Armillaria* resistance screening



- Initial inoculations on 1+0 stock, using birch stems as inoculum source
  - Year 1: grow 1+0
  - Year 2: inoculate, grow to 2+0
  - Year 3: plant in field-based trial
  - Years 4-6: Assess vigor (1-4), infection (Y/N), and mortality (Y/N).



# Fdi *Armillaria* resistance screening

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- 60 families/year
  - top 2 crosses/parent from pop being evaluated in the field
- 20 reps, 4 blocks, single tree plots
- Planted at ONE common location





# Fdi *Armillaria* resistance screening

- Populations inoculated:
  - 2021 inoculations, planted in 2022: NEH
  - 2022 inoculations, planted in 2023: NEL \*
  - 2023 inoculations, to plant in 2024: QL \*\*
  - 2024 inoculations, to plant in 2025: CP
  - Sowing in 2024, to inoculate in 2025: CT

\* Heat of 30 C in May 2023, at transplanting, could be reason of failed trial

\*\* To avoid potential heat stress at transplanting, putting trial in raised beds at KRS





# Fdi *Armillaria* resistance screening

- Plan to expand A.o. isolate collection:
  - Test pathogenicity of isolates
  - Test pathogenicity over time in subculture on artificial media
- Expand silver or paper birch bank at Simkinin or Bailey

# *Pissodes strobi* (Sx)

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- Moving on from raised beds with Sx for augmentation trials
- We have addressed media and historical patterns from old raised beds; however, allocated space for Fdi trial
- In 2023 we sowed a short-term trial for weevil screen – 140 families – to be planted in Skimikin in 2024



# *Pissodes strobi* (Sx)

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- Proposing a chemical ecology pilot study in partnership with PI at UBC/UNBC
- Test for host choice by weevil with referenced host genetic material -> testing for olfactory and physical cues that drive host selection. Keep source of weevil as a factor to test for differences in weevil structure





# *Dothistroma septosprum* (Pli)

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- Collected samples from Witset and Prince George in 2023
- Expanded our archive to 17 isolates
- Currently working on propagation and sporulation techniques
- Have shared isolated with UNBC





# Isolates from Pli needles



Identity	Count
<i>Dothistroma septosporum</i>	17
<i>Alternaria</i> spp.	1
<i>Cladosporium</i> spp.	10
<i>Epicoccum italicum</i>	2
<i>Genolevuria</i> spp.	3
<i>Hormonema macrosporum</i>	1
<i>Peniophora</i> spp.	2
<i>Pezizales</i> spp.	1
<i>Phaeotremella</i> spp.	1
<i>Pragmopora</i> spp.	1
<i>Pseudotremella</i> spp. / <i>Tremella</i> spp.	1
<i>Pseudozyma</i> spp.	1
<i>Ramularia</i> spp.	1
<i>Sydowia polyspora</i>	14
<i>Vishniacozyma victoriae</i>	1





Screening for White pine blister rust in Whitebark pine





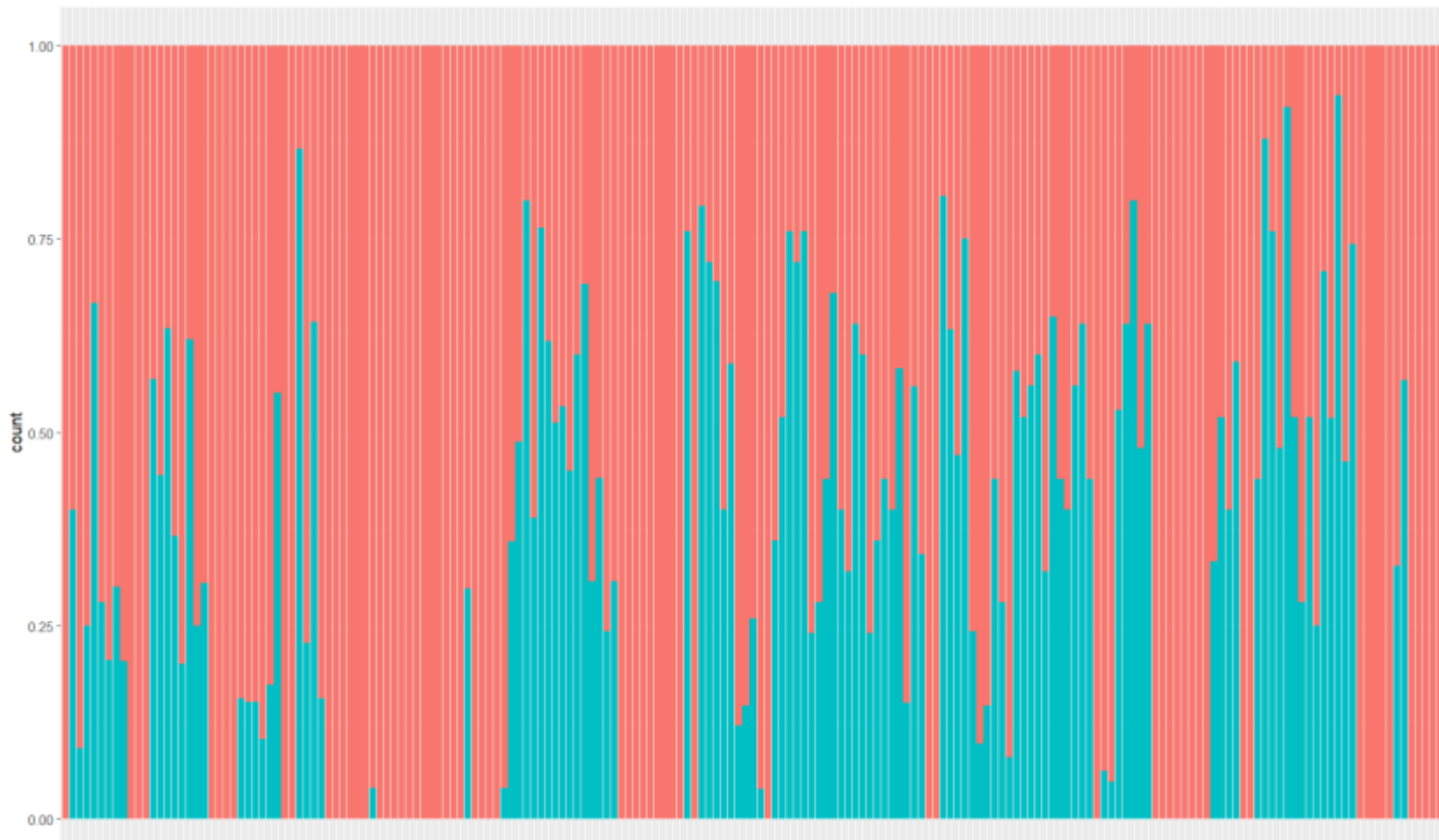


# Blister rust assessments

- Disease severity index:
  - Vigour (1-4)
  - # of cankers
  - Severity of cankers (1-9)
- Breeding values
  - Infection (Y/N)
  - Mortality(Y/N)



# Whitebark pine inoculations



- 168 families inoculated since 2021
- Will continue to inoculate 66 families/year
- 32 seedlings/fam, + 5 non inoculated controls
- Follow them up for 6 years in raised beds
- We observe 50% infection with our methods, robust measurement to screen

# In 2024:

- Continue with Fdi and armillaria screening
- Support field-based screening trials of Sx vs weevil
- Develop challenge methodology for Pli and Dothistroma
- Continue with Whitebark screening program vs WPBR
- Support field-based screening of Pli against Comandra rust
- Conduct a chemical ecology pilot on host selection cues by weevil on Sx
- Discuss and pave the way for an efficacy trial to control or suppress Phytophthora root rot in Pw
- Continue with lab and facilities upgrades needed for resistance programs
- Pending capacity, work on isolation and inoculation methodology for western gall rust

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